

CORRECTIONS

Proximity between Nucleotide/Dinucleotide and Metal Ion Binding Sites in DNA-Dependent RNA Polymerase from *Escherichia coli*, by Suresh C. Tyagi* and Felicia Y.-H. Wu, Volume 31, Number 28, July 21, 1992, pages 6447–6453.

Page 6447. Felicia Y.-H. Wu was omitted from the author byline. The byline should read as shown above. The address byline should read as follows: Department of Biochemistry and Cell Biology and Department of Pharmacological Sciences, State University of New York at Stony Brook, Stony Brook, New York, 11794-5215. The following permanent address for F.Y.-H.W. should be included: Division of Cancer Research, Institute of Biomedical Sciences, Academia Sinica, Taipei 11529, Taiwan, ROC.

Proton NMR Studies of Noncovalent Complexes of Cytochrome *c* Peroxidase–Cyanide with Horse and Yeast Ferri-cytochromes *c*, by Qian Yi, James E. Erman, and James D. Satterlee*, Volume 32, Number 41, October 19, 1993, pages 10988–10994.

Page 10990. In Figure 1, the placement of double bonds in the histidines shown in parts D and E is incorrect, and the ring positions are incorrectly labeled. To conform to IUPAC/IUB norms, the ring carbon labels 2 and 5 should be reversed. Then, the correct alignment of double bonds is along the N τ –C $_2$ bond and along the C $_4$ –C $_5$ bond. In part E the methionine is incorrectly labeled. The correct labeling is to change β to α and δ to β .

Complete Heme Proton Hyperfine Resonance Assignments of the *Glycera dibranchiata* Component IV Metcyano Monomer Hemoglobin, by Steve L. Alam and James D. Satterlee*, Volume 33, Number 13, April 5, 1994, pages 4008–4018.

Page 4011. The heme structure in Figure 1B is incorrectly labeled. The meso-position labels β and δ should be switched.

The double bonds in the histidine ring in Figure 1C are incorrectly positioned. The correct alignment of double bonds is along the N τ –C $_2$ bond and along the C $_4$ –C $_5$ bond.

Page 4013. In column 1, line 28, the text should be corrected to read ...substituent on heme pyrrole A, the....

Detection of Oligomeric and Monomeric Forms of P-glycoprotein in Multidrug Resistant Cells, by Marianne S. Poruchynsky and Victor Ling*, Volume 33, Number 14, April 12, 1994, pages 4163–4174.

Page 4166. Due to a printing error, the bands in Figure 2 are difficult to distinguish. An improved reproduction of the figure is shown below.

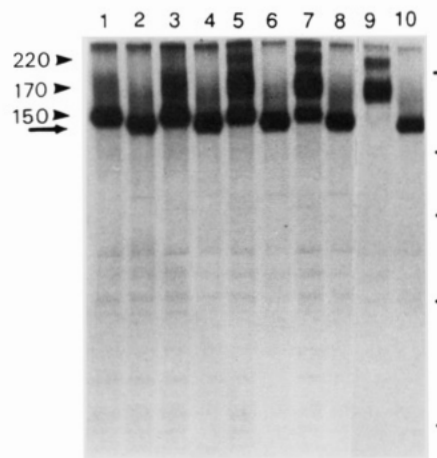


FIGURE 2: Pulse-chase labeling of P-glycoprotein in CHRC5 cells. Cells, which were either treated (lanes 2, 4, 6, 8, 10) or not treated (lanes 1, 3, 5, 7, 9) with 4 μ g/mL tunicamycin, were pulsed for 10 min with [35 S]methionine and chased in excess unlabeled methionine for 0, 5, 20, or 45 min or for 18 h, respectively. P-glycoprotein was immunoprecipitated from each sample using the monoclonal antibody C219, and separated by 7.5% SDS-PAGE. The positions of ~150-, 170-, and 220-kDa forms are indicated. The nonglycosylated ~140-kDa form of P-gp made in the presence of tunicamycin is indicated by the arrow.